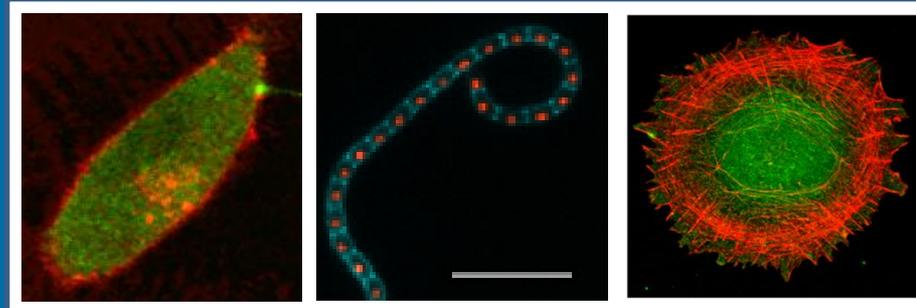




Uniwersytet  
Wrocławski



# BIO-IMAGIng in research INnovation and Education



**UNIVERSITY OF WROCLAW**

**Faculty of Biotechnology**

Tamka 2,

Przybyszewskiego 63/77,

WROCLAW, POLAND



- I. Confocal system (LSM510 META - Zeiss) - Laboratory of Cytobiochemistry**
- II. FACS - flow cytometer (Becton Dickinson FACSCalibur™ ) - Laboratory of Protein Biotechnology**
- III. Fluorescence microscopy system (AxioObserver" Z1 Cell Observer - Zeiss) - Laboratory of Molecular Microbiology**
- IV. Confocal system (FlowView 500 - Olympus) - Laboratory of Cell Pathology**

## Laboratory of Cytobiochemistry

Prof. Dr. hab. Aleksander Sikorski

➤ The system **is open to all interested users.**

**LSM510 META (Zeiss)** upgraded with FLIM/F(L)CS (TCSPC) Facility (PicoQuant)

### ➤ Features:

- META detector (412-748 nm)
- live cell imaging facility
- objectives: oil and water immersed (40x, 63x), dry (10x, 20x)
- two SPADs (possible FCCS measurements)
- filters for FLIM/F(L)CS
- laser lines [nm]: cw: 405, 454, 488, 514, 561, 633;  
pulsed diode lasers: 470 (FWHM ~58ps), 635 (FWHM: ~59ps)  
repetition frequency 31,25kHz – 80 MHz.

### ➤ Softwares:

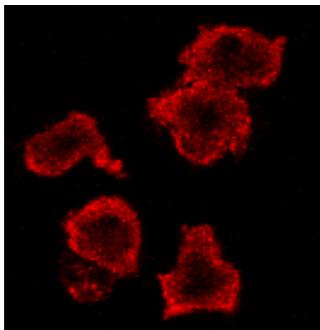
- **ZEN** - support microscope operation and image analysis: FRAP, FRET, deconvolution, 3D projections, linear unmixing, colocalization, time lapse imaging,
- **SymPhoTime** – operation and analysis of time-resolved imaging/measurements:  
F(L)CS fitting to four models implemented: pure diffusions, triplet, conformational and protonation.



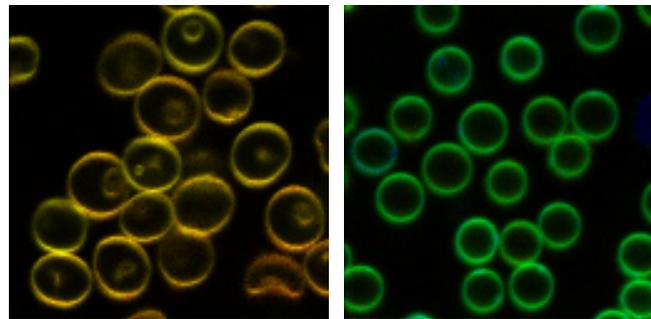


➤ **Research focus and imaging techniques:**

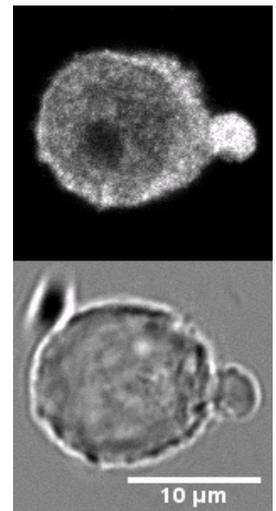
- imaging studies of fixed specimen,
- determination of fluorescent lipid diffusion in liposomes by **FRAP**,
- **FLIM** in lipid rafts imaging
- determination of interaction between eGFP-tagged proteins and fluorescently labeled,
- plasma membrane by acceptor bleaching **FRET** and **FLIM-FRET**,
- spectrin engagement in immunological synapse formation,
- determination of PKB activity in living cells with FRET based probe.



**Spectrin in HL-60  
during apoptosis**



**Di-4-ANEPPDHQ in human RBC  
(time-resolved image)**



# FACS - flow cytometer

## Laboratory of Protein Biotechnology

Dr hab. Ewa Marcinkowska

➤ The BD FACSCalibur flow cytometer was purchased by Wrocław Research Centre EIT+ as an equipment necessary to complete the projects covered by the programme BioMed (access restricted).

### ➤ features:

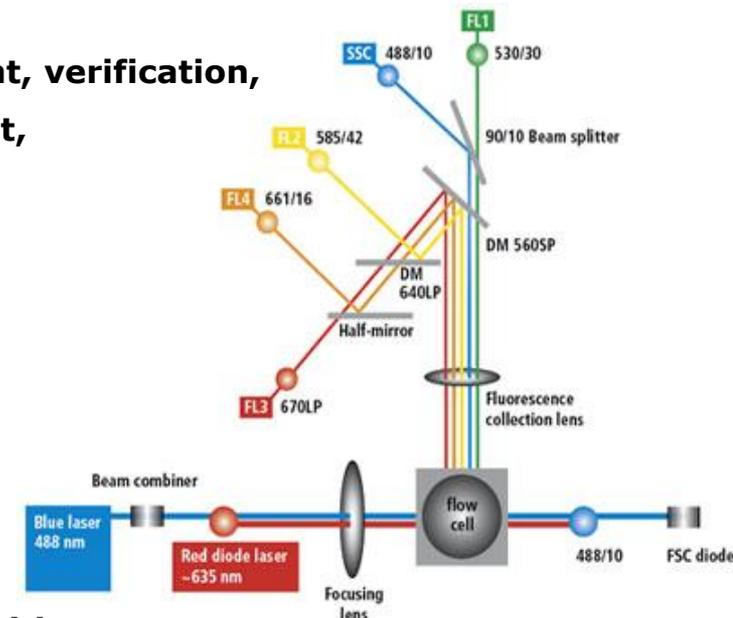
- a multiparameter system
- system for routine cell analysis, assay development, verification, and identification of cellular populations of interest,
- dual-laser technology,
- 4-colour analysis capabilities.

### ➤ planned equipment base

- separate equipment for efficient and precise cell sorting in sterile conditions,
- separate for detection of up to 18 colors simultaneously with 4 lasers and digital data acquisition system.



**Becton Dickinson FACSCalibur™**



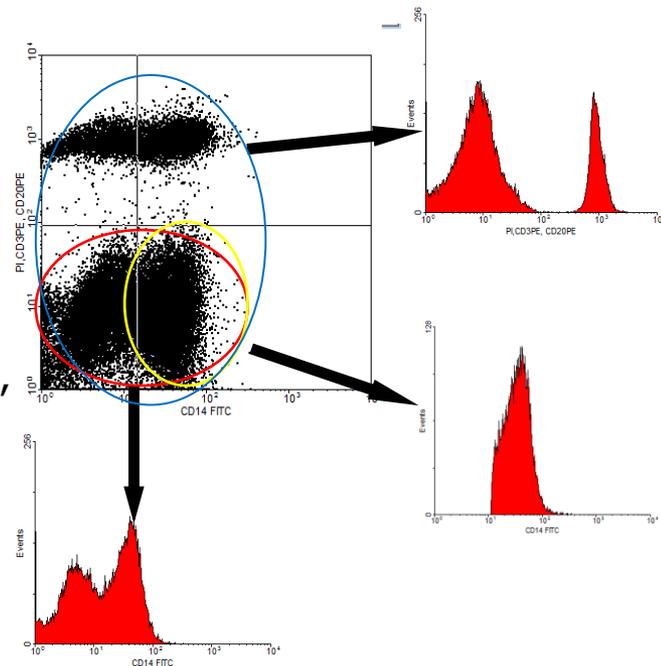
## ➤ Research focus and imaging techniques:

### Projects:

- "Identification of molecular markers of diseases related to nuclear receptors dysfunction and implementation in diagnostics."
- "Synthesis and characterization of bionanomaterials for specific medical applications."
- "Employment of ferrum and haem assimilation by *Porphyromonas gingivalis* bacteria in the prevention and treatment of periodontopathy."

### Measurements :

- of cell surface antigens expression,
- of expression levels of intracellular proteins,
- of cytokine levels in blood serum
- of transfection efficiency using fluorescent proteins,
- detection of apoptosis using various techniques,
- cell cycle analysis using measurements of DNA content.



# Fluorescence Microscopy Systems

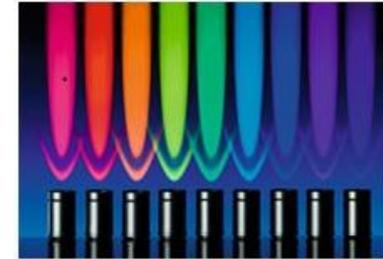
## Laboratory of Molecular Microbiology

**Prof. Dr. hab. Jolanta Zakrzewska-Czerwińska**

The microscope was purchased by Wrocław Research Centre EIT+ as an equipment necessary to complete the projects covered by the programme BioMed (access restricted).

### ➤ **Features:**

- fully automated,
- BF, Fluorescence, Phase Contrast and DIC,
- filter sets (DAPI (49) , TexasRed (43 HE), GFP (38 HE), Cy 5 (50))
- COLIBRI system,
- dry and oil objectives (10x, 40x, 100x),
- combined with Definite focus (eliminate Z drift),
- modules multichannel, Z-Stack, time lapse, autofocus, mark & find, mosaiX,
- temperature, CO<sub>2</sub> control,
- AxioVision - image analysis software for acquisition and image processing.



**AxioObserver" Z1 Cell Observer  
(Zeiss)**

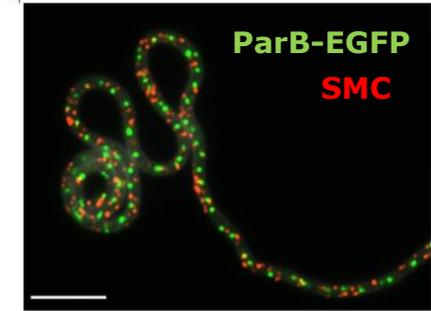
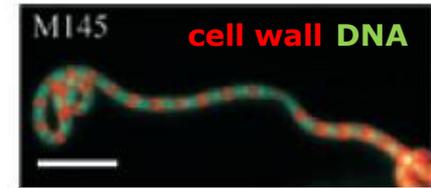
### ➤ **planned equipment base**

- more advanced software for image analysis, system for FRAP.

➤ **Research focus and imaging techniques:**

**Projects:**

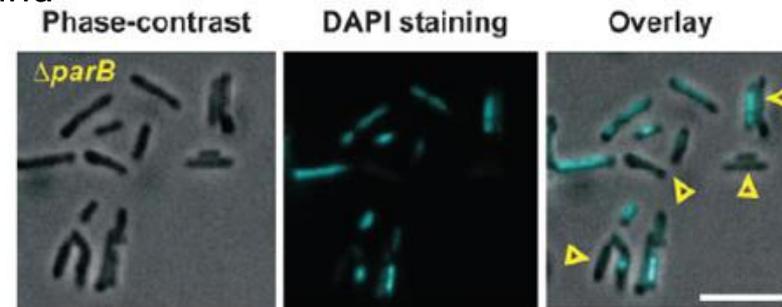
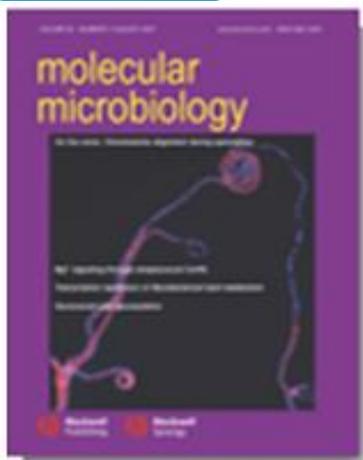
- Chromosome replication, condensation and segregation in *Streptomyces* and *Mycobacterium*,
- Development of *Streptomyces venezuelae*,
- Genome mining for drug discovery regulation of chromosome replication in *Streptomyces*.



*Streptomyces coelicolor*

**Techniques:**

- localization of proteins and chromosomal regions (gfp, cherry fusion protein)
- observation of migration of proteins and chromosomes in **real time**
- immunostaining.



*Mycobacterium smegmatis*

## Laboratory of Protein Engineering

Dr hab. Artur Krężel

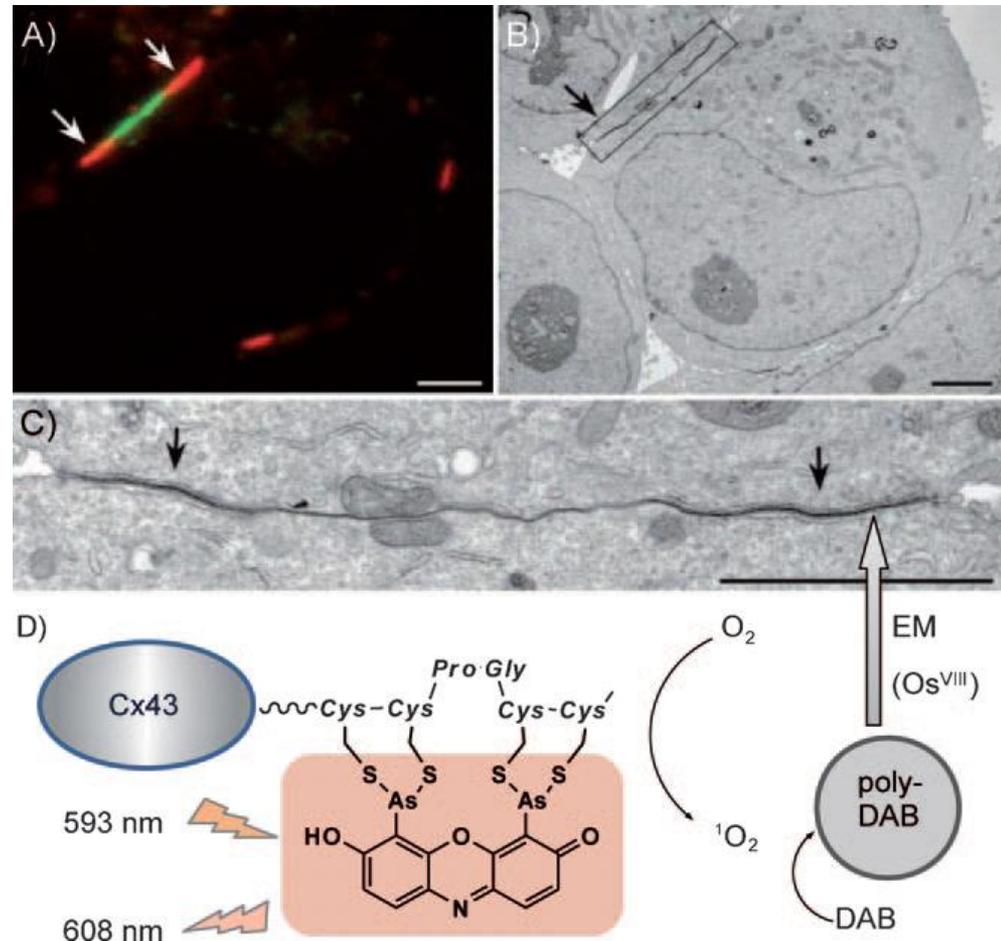
### ➤ Innovative imaging techniques

#### Biarsenical fluorescent tags

Two components:

- very short (6- to 12-residue), tetracysteine sequence (CCXXCC), placed at a protein terminus;
- nonluminescent biarsenical probe;

Protein contains the tetracysteine motif becomes highly fluorescent upon labeling with a nonluminescent biarsenical probe, and forms very stable covalent complexes.



## Laboratory Of Cell Pathology

**Prof. Dr. hab. Maria Malicka-Błaszkiwicz**

➤ **The system is open to all interested users.**

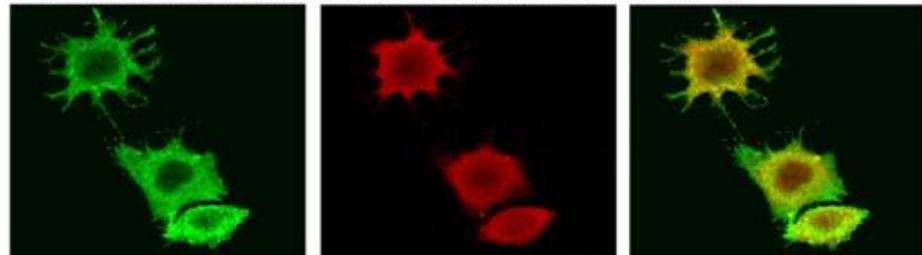
➤ **Equipment:**

Confocal fluorescence microscope FlowView 500 Olympus

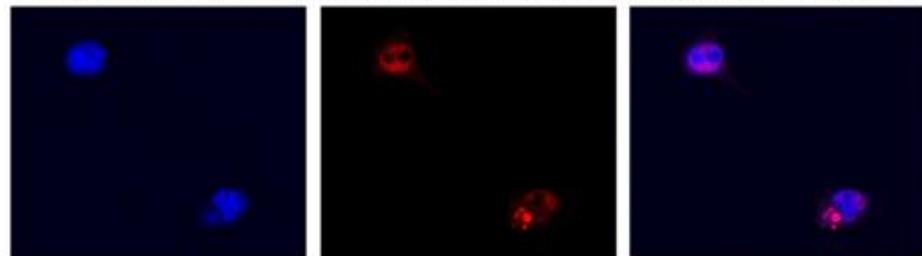
➤ **Research interest**

- The role of actin and actin binding proteins (ABPs) in the cell pathology and clinic;
- Drug induced apoptosis as a therapeutic tool;
- Cell culture as a model in cytotoxicity assay and biomedical studies.

NRK treated with 10M MTX for 48h  
stained with anti  $\beta$ -actin Ab (green) and rhodamine-phalloidin (red)



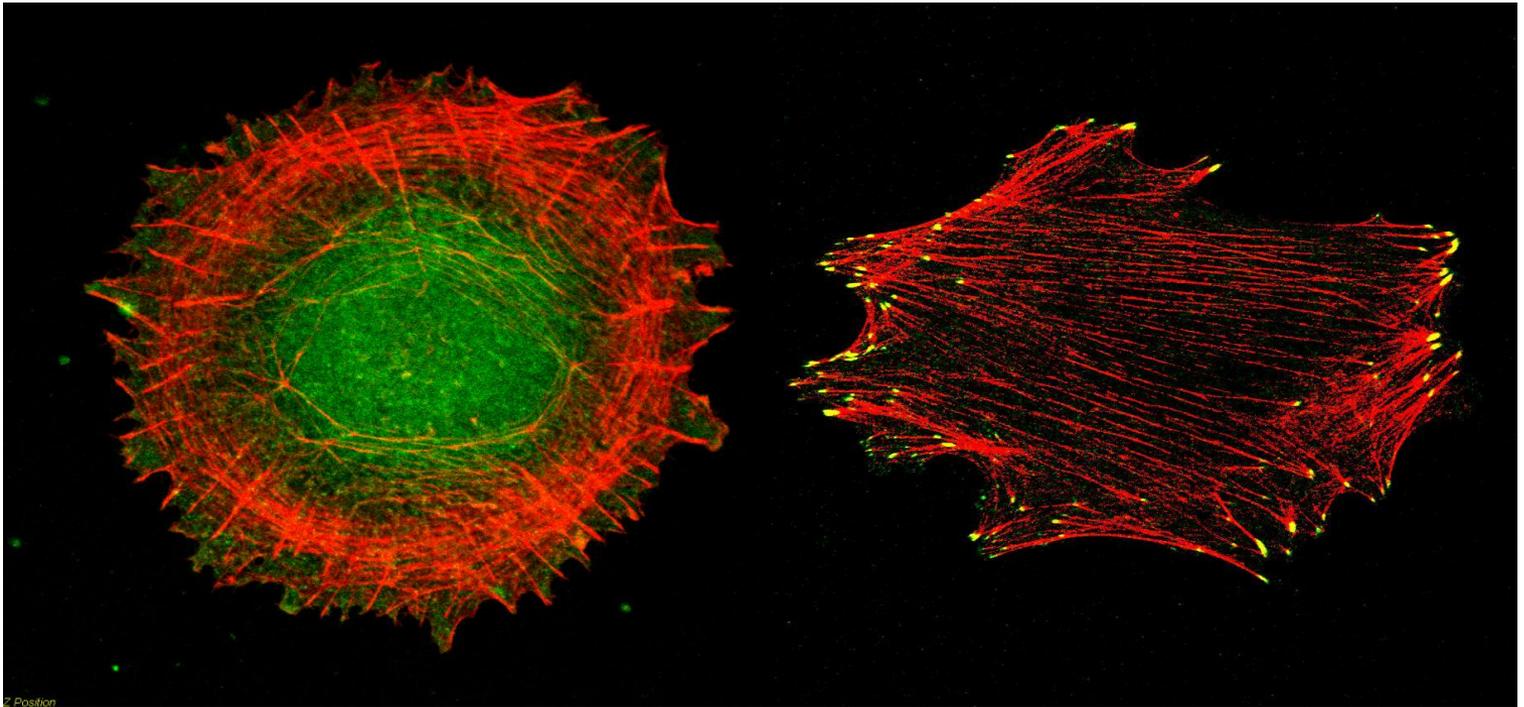
stained with anti active caspase 3 Ab (red), annexin V-Fluos (green), chromomycin





**A 375 melanoma cell**

**Fibroblast**



**green – gelsolin**

**green – vinculin**

**red – rhodamine- phalloidin**