Errata do rozprawy doktorskiej pt.
"Rola białka ATRX w organizacji chromatyny neuronów hipokampa mózgu szczura" autorstwa Dagmary Holm-Kaczmarek w odpowiedzi na recenzję dr Agnieszki Girstun

8. Bibliografia

Abidi, F. E., Cardoso, C., Lossi, A. M., Lowry, R. B., Depetris, D., Mattéi, M. G., Lubs, H. A., Stevenson, R. E., Fontes, M., Chudley, A. E., & Schwartz, C. E. (2005). Mutation in the 5′ alternatively spliced region of the XNP/ATR-X gene causes Chudley-Lowry syndrome. *European Journal of Human Genetics*, *13*(2), 176–183. https://doi.org/10.1038/sj.ejhg.5201303

Aguilera, P. & López-Contreras, A. J. ATRX, a guardian of chromatin. (2023). *Trends Genet.* 39(6):505-519. doi: 10.1016/j.tig.2023.02.009. Epub 2023 Mar 7. PMID: 36894374.

Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2008). Molecular Biology of the Cell (5th ed.). New York: *Garland Science.*

Alcamí, P. & Pereda, A. E. (2019). Beyond plasticity: the dynamic impact of electrical synapses on neural circuits. *Nature Reviews Neuroscience*, *20*(5), 253–271. https://doi.org/10.1038/s41583-019-0133-5

Allen, K. D., Gourov, A. V., Harte, C., Gao, P., Lee, C., Sylvain, D., Splett, J. M., Oxberry, W. C., van de Nes, P. S., Troy-Regier, M. J., Wolk, J., Alarcon, J. M., & Hernández, A. I. (2014). Nucleolar Integrity Is Required for the Maintenance of Long-Term Synaptic Plasticity. *PLoS ONE*, *9*(8), e104364. https://doi.org/10.1371/journal.pone.0104364

Argilli, E., Sibley, D.R., Malenka, R.C., England, P.M., Bonci, A. (2008). Mechanism and time course of cocaine-induced long-term potentiation in the ventral tegmental area. *J Neurosci.* 2008 Sep 10;28(37):9092-100. doi: 10.1523/JNEUROSCI.1001-08.2008. PMID: 18784289; PMCID: PMC2586328.

Banker, G.A. & Cowan, W.M. (1977) Rat hippocampal neurons in dispersed cell culture. *Brain Res*. 1977 May 13;126(3):397-42. doi: 10.1016/0006-8993(77)90594-7. PMID: 861729.

Barr, M. & Bertram, E. A Morphological Distinction between Neurones of the Male and Female, and the Behaviour of the Nucleolar Satellite during Accelerated Nucleoprotein Synthesis. (1949). *Nature* 163, 676–677. https://doi.org/10.1038/163676a0

Baumann, C., & De La Fuente, R. (2009). ATRX marks the inactive X chromosome (Xi) in somatic cells and during imprinted X chromosome inactivation in trophoblast stem cells. *Chromosoma*, *118*(2), 209–222. https://doi.org/10.1007/s00412-008-0189-x

Beaudoin, G. M. 3rd, Seung-Hye L., Dipika, S., Yang, Y., Yu-Gie, N., Louis, F. R., Jyothi, A. (2012) Culturing pyramidal neurons from the early postnatal mouse hippocampus and cortex. *Nat Protoc*. 2012 Sep;7(9):1741-54. doi: 10.1038/nprot.2012.099. Epub 2012 Aug 30. PMID: 22936216.

Benito, E., & Barco, A. (2015). The neuronal activity-driven transcriptome. *Molecular neurobiology*, *51*(3), 1071–1088. https://doi.org/10.1007/S12035-014-8772-Z

Bérubé, N. G., Mangelsdorf, M., Jagla, M., Vanderluit, J., Garrick, D., Gibbons, R. J., Higgs, D. R., Slack, R. S., & Picketts, D. J. (2005). The chromatin-remodeling protein ATRX is critical for neuronal survival during corticogenesis. *Journal of Clinical Investigation*, *115*(2), 258–267. https://doi.org/10.1172/JCI200522329

Bérubé, N. G., Smeenk, C. A., & Picketts, D. J. (2000). Cell cycle-dependent phosphorylation of the ATRX protein correlates with changes in nuclear matrix and chromatin association. *Human Molecular Genetics*, *9*(4), 539–547. https://doi.org/10.1093/hmg/9.4.539

Biała, G. (2007). Pamięć a uzależnienia lekowe : rola kalcyneuryny i hipokampa. *Postepy Hig Med Dosw (online)*, *61*, 199–203. http://www.phmd.pl/pub/phmd/vol_61/10358.pdf

Billia, F., Baskys, A., Carlen, P.L., De Boni, U. (1992). Rearrangement of centromeric satellite DNA in hippocampal neurons exhibiting long-term potentiation. *Molecular Brain Research.* Volume 14, Issues 1-2. https://doi.org/10.1016/0169-328X(92)90016-5.

Birnstiel, M. L. & Hyde, B. B. (1963). Protein synthesis by isolated pea nucleoli. *The Journal of cell biology*, *18*(18), 41–50. https://doi.org/10.1083/jcb.18.1.41

Bliss, T.V. & Lomo, T. (1973). Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol.* Jul;232(2):331-56. doi: 10.1113/jphysiol.1973.sp010273.

Bokota, G., Sroka, J., Basu, S., Das, N., Trzaskoma, P., Yushkevich, Y., Grabowska, A., Magalska, A., & Plewczynski, D. (2021). PartSeg: a tool for quantitative feature extraction from 3D microscopy images for dummies. *BMC Bioinformatics*, *22*(1), 1–19. https://doi.org/10.1186/s12859-021-03984-1

Botchkarev, V. A., Gdula, M. R., Mardaryev, A. N., Sharov, A. A., & Fessing, M. Y. (2012). Epigenetic regulation of gene expression in keratinocytes. *Journal of Investigative Dermatology*, *132*(11), 2505–2521. https://doi.org/10.1038/jid.2012.182

Boulon, S., Westman, B. J., Hutten, S., Boisvert, F. M., & Lamond, A. I. (2010). The Nucleolus under Stress. W *Molecular Cell* (T. 40, Numer 2, ss. 216–227). Mol Cell. https://doi.org/10.1016/j.molcel.2010.09.024

Bramham, C. R. (2010). LTP ≠ learning: Lessons from short-term plasticity. *Frontiers in Behavioral Neuroscience*, *4*(FEB), 3–4. https://doi.org/10.3389/neuro.08.003.2010

Brangwynne, C. P., Mitchison, T. J., & Hyman, A. A. (2011). Active liquid-like behavior of nucleoli determines their size and shape in Xenopus laevis oocytes. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(11), 4334–4339. https://doi.org/10.1073/pnas.1017150108

Bunina, D., Abazova, N., Diaz, N., Noh, K. M., Krijgsveld, J., & Zaugg, J. B. (2020). Genomic Rewiring of SOX2 Chromatin Interaction Network during Differentiation of ESCs to Postmitotic Neurons. *Cell Systems*, *10*(6), 480-494.e8. https://doi.org/10.1016/j.cels.2020.05.003

Burke, B., & Stewart, C. L. (2013). The nuclear lamins: Flexibility in function. *Nature Reviews Molecular Cell Biology*, *14*(1), 13–24. https://doi.org/10.1038/nrm3488

Byrne, M.J., Putkey, J.A., Neal Waxham, M. et al. (2009). Dissecting cooperative calmodulin binding to CaM kinase II: a detailed stochastic model. J Comput Neurosci 27, 621–638. https://doi.org/10.1007/s10827-009-0173-3

Cabral JM, Oh HS, Knipe DM. ATRX promotes maintenance of herpes simplex virus heterochromatin during chromatin stress. (2018). *Elife*. 2018 Nov 22;7:e40228. doi: 10.7554/eLife.40228. PMID:

30465651; PMCID: PMC6307862.

Chaurasia, A., Park, S.H., Seo, J.W., Park, C.K. (2016). Immunohistochemical Analysis of ATRX, IDH1 and p53 in Glioblastoma and Their Correlations with Patient Survival. *J Korean Med Sci.* 2016 Aug;31(8):1208-14. doi: 10.3346/jkms.2016.31.8.1208. Epub 2016 May 30. PMID: 27478330; PMCID: PMC4951549.

Cheung, T. H. C., & Cardinal, R. N. (2005). Hippocampal lesions facilitate instrumental learning with delayed reinforcement but induce impulsive choice in rats. *BMC Neuroscience*, *6*, 1–24. https://doi.org/10.1186/1471-2202-6-36

Clynes, D., Higgs, D. R., & Gibbons, R. J. (2013). The chromatin remodeller ATRX: A repeat offender in human disease. *Trends in Biochemical Sciences*, *38*(9), 461–466. https://doi.org/10.1016/j.tibs.2013.06.011

Clynes, D., Jelinska, C., Xella, B., Ayyub, H., Taylor, S., Mitson, M., Bachrati, CZ., Higgs, D. R., Gibbons, R. J. (2014). ATRX dysfunction induces replication defects in primary mouse cells. *PLoS One.* 2014 Mar 20;9(3):e92915. doi: 10.1371/journal.pone.0092915. PMID: 24651726; PMCID: PMC3961441.

Cremer, T., Cremer, M., Hübner, B., Strickfaden, H., Smeets, D., Popken, J., Sterr, M., Markaki, Y., Rippe, K., Cremer, C. (2015). The 4D nucleome: Evidence for a dynamic nuclear landscape based on co-aligned active and inactive nuclear compartments. *FEBS Lett*. 7;589(20 Pt A):2931-43. doi: 10.1016/j.febslet.2015.05.037. Epub 2015 May 28. PMID: 26028501.

Dai, M.-S., Zeng, S. X., Jin, Y., Sun, X.-X., David, L., & Lu, H. (2004). Ribosomal Protein L23 Activates p53 by Inhibiting MDM2 Function in Response to Ribosomal Perturbation but Not to Translation Inhibition. *Molecular and Cellular Biology*, *24*(17), 7654–7668. https://doi.org/10.1128/mcb.24.17.7654-7668.2004

De Carlos, J. A., & Borrell, J. (2007). A historical reflection of the contributions of Cajal and Golgi to the foundations of neuroscience. *Brain Research Reviews*, *55*(1), 8–16. https://doi.org/10.1016/j.brainresrev.2007.03.010

De Castro, F., López-Mascaraque, L., & De Carlos, J. A. (2007). Cajal: Lessons on brain development. *Brain Research Reviews*, *55*(2 SPEC. ISS.), 481–489. https://doi.org/10.1016/j.brainresrev.2007.01.011

Dekker, J., Rippe, K., Dekker, M., Kleckner, N.. Capturing chromosome conformation. (2002). *Science.* Feb 15;295(5558):1306-11. doi: 10.1126/science.1067799. PMID: 11847345.

Desnoyers, S., Kaufmann, S. H., & Poirier, G. G. (1996). Alteration of the nucleolar localization of poly(ADP-ribose) polymerase upon treatment with transcription inhibitors. *Experimental Cell Research*, *227*(1), 146–153. https://doi.org/10.1006/excr.1996.0259

Duvernoy, H. M. (2005). The Human Hippocampus: Functional Anatomy, Vascularization and Serial Sections. *Springer*. ISBN: 978-3540209817.

Dyer, M. A., Qadeer, Z. A., Valle-Garcia, D., & Bernstein, E. (2017). ATRX and DAXX: Mechanisms and mutations. *Cold Spring Harbor Perspectives in Medicine*, *7*(3), 1–16. https://doi.org/10.1101/cshperspect.a026567

Eiland, L., Ramroop, J., Hill, M. N., Manley, J., McEwen, B. S. (2012). Chronic juvenile stress produces corticolimbic dendritic architectural remodeling and modulates emotional behavior in male and female rats. *Psychoneuroendocrinology*. Jan;37(1):39-47. doi: 10.1016/j.psyneuen.2011.04.015.

Elsaesser, S. J., Goldberg, A. D., & Allis, C. D. (2011). *Elsaesser S J (2010) New functions for an old variant no substitute for histone H3.3*. *20*(2), 110–117. https://doi.org/10.1016/j.gde.2010.01.003

Felsenfeld, G. & Groudine, M.. Controlling the double helix. (2003). *Nature*. Jan 23;421(6921):448-53. doi: 10.1038/nature01411. PMID: 12540921.

Feric, M., Vaidya, N., Harmon, T. S., Mitrea, D. M., Zhu, L., Richardson, T. M., Kriwacki, R. W., Pappu, R. V., & Brangwynne, C. P. (2016). Coexisting Liquid Phases Underlie Nucleolar Subcompartments. *Cell*, *165*(7), 1686–1697. https://doi.org/10.1016/j.cell.2016.04.047

Fussner, E., Strauss, M., Djuric, U., Li, R., Ahmed, K., Hart, M., Ellis, J., & Bazett-Jones, D. P. (2012). Open and closed domains in the mouse genome are configured as 10-nm chromatin fibres. *EMBO Reports*, *13*(11), 992–996. https://doi.org/10.1038/embor.2012.139

Freed, E.F., Bleichert, F., Dutca, L.M., Baserga, S.J. (2010). When ribosomes go bad: diseases of ribosome biogenesis. *Mol Biosyst*. 2010 Mar;6(3):481-93. doi: 10.1039/b919670f. Epub 2010 Jan 11. PMID: 20174677; PMCID: PMC2965583.

Galganski, L., Urbanek, M. O., & Krzyzosiak, W. J. (2017). Nuclear speckles: Molecular organization, biological function and role in disease. *Nucleic Acids Research*, *45*(18), 10350–10368. https://doi.org/10.1093/nar/gkx759

Gapp, K., Jawaid, A., Sarkies, P., Bohacek, J., Pelczar, P., Farinelli, L., Miska, E., & Mansuy, I. M. (2014). Feeding the Next Generation of Corn Ethanol Byproducts To. *Nature Neuroscience*, *17*(5), 667–669. https://doi.org/10.1038/nn.3695.Implication

Garrick, D., Samara, V., McDowell, T. L., Smith, A. J. H., Dobbie, L., Higgs, D. R., & Gibbons, R. J. (2004). A conserved truncated isoform of the ATR-X syndrome protein lacking the SWI/SNF-homology domain. *Gene*, *326*(1–2), 23–34. https://doi.org/10.1016/j.gene.2003.10.026

Gibbons, R. J., & Higgs, D. R. (2000). *Molecular – Clinical Spectrum of the ATR-X Syndrome*. *212*, 204–212.

Gibbons, R. J., Wilkie, A. O. M., Weatherall, D. J., & Higgs, D. R. (1991). A newly defined X linked mental retardation syndrome associated with α thalassaemia. *Journal of Medical Genetics*, *28*(11), 729–733. https://doi.org/10.1136/jmg.28.11.729

Gluck, M. A., & Myers, C. E. (1997). Psychobiological models of hippocampal function in learning and memory. *Annual Review of Psychology*, *48*, 481–514. https://doi.org/10.1146/annurev.psych.48.1.481

Goldberg, A. D., Banaszynski, L. A., Noh, K., Lewis, P. W., Elsaesser, J., Stadler, S., Dewell, S., Law, M., Guo, X., Li, X., Wen, D., Chapgier, A., Dekelver, R. C., Miller, J. C., Boydston, E. A., Holmes, M. C., Gregory, P. D., John, M., Rafii, S., Urnov, F. D., Zheng D., Allis C. D. (2011). Distinct Factors Control Histone Variant H3.3 Localization at Specific Genomic Regions. *Cell*. *140*(5), 678–691. https://doi.org/10.1016/j.cell.2010.01.003.

Gorlewicz, A., Krawczyk, K., Szczepankiewicz, A. A., Trzaskoma, P., Mulle, C., & Wilczynski, G. M. (2020). Colocalization Colormap -an ImageJ Plugin for the Quantification and Visualization of Colocalized Signals. *Neuroinformatics*, *18*(4), 661–664. https://doi.org/10.1007/s12021-020-09465-9

Grabowska, A., Sas-Nowosielska, H., Wojtas, B., Holm-Kaczmarek, D., Januszewicz, E., Yushkevich, Y., Czaban, I., Trzaskoma, P., Krawczyk, K., Gielniewski, B., Martin-Gonzalez, A., Filipkowski, R. K., Olszynski, K. H., Bernas, T., Szczepankiewicz, A. A., Sliwinska, M. A., Kanhema, T., Bramham, C. R., Bokota, G., Plewczynski, D., Wilczynski, G. M., Magalska, A. (2022).

Activation-induced chromatin reorganization in neurons depends on HDAC1 activity. *Cell Rep*. 2022 Feb 15;38(7):110352. doi: 10.1016/j.celrep.2022.110352.

Grummt, I., & Ladurner, A. G. (2008). A Metabolic Throttle Regulates the Epigenetic State of rDNA. W *Cell*. https://doi.org/10.1016/j.cell.2008.04.026

Gugustea, R., Tamming, R. J., Martin-Kenny, N., Bérubé, N. G., & Leung, L. S. (2020). Inactivation of ATRX in forebrain excitatory neurons affects hippocampal synaptic plasticity. *Hippocampus*, *30*(6), 565–581. https://doi.org/10.1002/hipo.23174

Hashimoto, H., Vertino, P. M., & Cheng, X. (2010). Molecular coupling of DNA methylation and histone methylation. *Epigenomics*, *2*(5), 657–669. https://doi.org/10.2217/epi.10.44

Hergeth, S. P., Schneider, R. (2015). The H1 linker histones: multifunctional proteins beyond the nucleosomal core particle. *EMBO Rep*. 2015 Nov;16(11):1439-53. doi: 10.15252/embr.201540749. Epub 2015 Oct 15. PMID: 26474902; PMCID: PMC4641498.

Herre, M., & Korb, E. (2019). The chromatin landscape of neuronal plasticity. *Current Opinion in Neurobiology*, *59*, 79–86. https://doi.org/10.1016/j.conb.2019.04.006

Hetman, M., & Pietrzak, M. (2012). Emerging roles of the neuronal nucleolus. *Trends in Neurosciences*, *35*(5), 305–314. https://doi.org/10.1016/j.tins.2012.01.002

Hetman, M., & Slomnicki, L. P. (2019). Ribosomal biogenesis as an emerging target of neurodevelopmental pathologies. *Journal of Neurochemistry*, *148*(3), 325–347. https://doi.org/10.1111/jnc.14576

Higgs, D. R., Garrick, D., Anguita, E., De Gobbi, M., Hughes, J., Muers, M., Vernimmen, D., Lower, K., Law, M., Argentaro, A., Deville, M. A., Gibbons, R. (2005). Understanding alpha-globin gene regulation: Aiming to improve the management of thalassemia. *Ann N Y Acad Sci*. 2005;1054:92-102. doi: 10.1196/annals.1345.012. PMID: 16339655.

Horch, H. W., & Katz, L. C. (2002). BDNF release from single cells elicits local dendritic growth in nearby neurons. *Nature Neuroscience*, *5*(11), 1177–1184. https://doi.org/10.1038/nn927

Ishov, A. M., Sotnikov, A. G., Negorev, D., Vladimirova, O. V., Neff, N., Kamitani, T., Yeh, E. T. H., Strauss, J. F., & Maul, G. G. (1999). PML is critical for ND10 formation and recruits the PML-interacting protein Daxx to this nuclear structure when modified by SUMO-1. *Journal of Cell Biology*, *147*(2), 221–233. https://doi.org/10.1083/jcb.147.2.221

James, A., Wang, Y., Raje, H., Rosby, R., & DiMario, P. (2014). Nucleolar stress with and without p53. *Nucleus*, *5*(5), 402–426. https://doi.org/10.4161/nucl.32235

Janiszewski, L., Barbacka-Surowiak, G., Surowiak, J. (1993). Neurofizjologia porównawcza. Warszawa: *Wydawnictwo Naukowe PWN*.

Jaworski, J., Kalita, K., Knapska, E. (2018) c-Fos and neuronal plasticity: the aftermath of Kaczmarek's theory. *Acta Neurobiol Exp (Wars)*. 2018;78(4):287-296. PMID: 30624427.

Jin, B., & Robertson, K. D. (2013). DNA methyltransferases, DNA damage repair, and cancer. *Advances in Experimental Medicine and Biology*, *754*, 3–29. https://doi.org/10.1007/978-1-4419-9967-2_1

Kaczmarek, L. (1992). Expression of c-fos and other genes encoding transcription factors in long-term potentiation. *Behavioral and Neural Biology*, *57*(3), 263–266. https://doi.org/10.1016/0163-1047(92)90276-A

Kaczmarek, L. & Chaudhuri, A. (1997). Sensory regulation of immediate-early gene expression in mammalian visual cortex: implications for functional mapping and neural plasticity. *Brain Res Brain Res Rev*. 1997 Apr;23(3):237-56. doi: 10.1016/s0165-0173(97)00005-2. PMID: 9164673.

Kalita, K., Makonchuk, D., Gomes, C., Zheng, J. J., & Hetman, M. (2008). Inhibition of nucleolar transcription as a trigger for neuronal apoptosis. *Journal of Neurochemistry*, *105*(6), 2286–2299. https://doi.org/10.1111/j.1471-4159.2008.05316.x

Kaufmann, W. E., & Moser, H. W. (2000). Dendritic anomalies in disorders associated with mental retardation. *Cerebral Cortex*, *10*(10), 981–991. https://doi.org/10.1093/cercor/10.10.981

Kernohan, K. D., Vernimmen, D., Gloor, G. B., & Bérubé, N. G. (2014). Analysis of neonatal brain lacking ATRX or MeCP2 reveals changes in nucleosome density, CTCF binding and chromatin looping. *Nucleic Acids Research*, *42*(13), 8356–8368. https://doi.org/10.1093/nar/gku564

Khatau, S. B., Hale, C. M., Stewart-Hutchinson, P. J., Patel, M. S., Stewart, C. L., Searson, P. C., Hodzic, D., & Wirtz, D. (2009). A perinuclear actin cap regulates nuclear shape. *Proceedings of the National Academy of Sciences of the United States of America*, *106*(45), 19017–19022. https://doi.org/10.1073/pnas.0908686106

Kim, S., Yu, N. K., & Kaang, B. K. (2015). CTCF as a multifunctional protein in genome regulation and gene expression. *Experimental and Molecular Medicine*, *47*(6). https://doi.org/10.1038/EMM.2015.33

Kobayashi, T. (2008). A new role of the rDNA and nucleolus in the nucleus - RDNA instability maintains genome integrity. *BioEssays*, *30*(3), 267–272. https://doi.org/10.1002/bies.20723

Konturek, S. (1993). Fizjologia człowieka. Tom IV - Neurofizjologia. *Wydawnictwo Lekarskie PZWL.* Warszawa.

Konorski, J. (1948). Conditioned reflexes and neuron organization. *Cambridge University Press.*

Kornberg, R. (1974). Chromatin Structure : A Repeating Unit of Histones and DNA Chromatin structure is based on a repeating unit of eight. *Science*, *184*, 868–871.

Kossut, M. (2019). Basic mechanism of neuroplasticity. *Neuropsychiatria i Neuropsychologia*. 14(1), 1-8. https://doi.org/10.5114/nan.2019.87727

Kouzarides, T. (2007). Chromatin Modifications and Their Function. *Cell*, *128*(4), 693–705. https://doi.org/10.1016/j.cell.2007.02.005

Lafontaine, D. L. J., Riback, J. A., Bascetin, R., & Brangwynne, C. P. (2020). The nucleolus as a multiphase liquid condensate. W *Nature Reviews Molecular Cell Biology* (T. 22, Numer 3, ss. 165–182). Nature Research. https://doi.org/10.1038/s41580-020-0272-6

Lewis, P. W., Elsaesser, S. J., Noh, K. M., Stadler, S. C., & Allis, C. D. (2010). Daxx is an H3.3-specific histone chaperone and cooperates with ATRX in replication-independent chromatin assembly at telomeres. *Proceedings of the National Academy of Sciences of the United States of America*, *107*(32), 14075–14080. https://doi.org/10.1073/pnas.1008850107

Lindström, M. S., Jurada, D., Bursac, S., Orsolic, I., Bartek, J., & Volarevic, S. (2018). Nucleolus as an emerging hub in maintenance of genome stability and cancer pathogenesis. *Oncogene*, *37*(18), 2351–2366. https://doi.org/10.1038/s41388-017-0121-z

Liu, Y., Liang, S., & Tartakoff, A. M. (1996). Heat shock disassembles the nucleolus and inhibits nuclear protein import and poly(A)+ RNA export. *EMBO Journal*, *15*(23), 6750–6757. https://doi.org/10.1002/j.1460-2075.1996.tb01064.

Longstaff A. (2011) Krótkie wykłady. Neurobiologia. Warszawa: *Wydawnictwo Naukowe PWN.*

Luger, K., Mäder, A. W., Richmond, R. K., Sargent, D. F., & Richmond, T. J. (1997). Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature*, *389*(6648), 251–260. https://doi.org/10.1038/38444

Maggi, L. B., Kuchenruether, M., Dadey, D. Y. A., Schwope, R. M., Grisendi, S., Townsend, R. R., Pandolfi, P. P., & Weber, J. D. (2008). Nucleophosmin Serves as a Rate-Limiting Nuclear Export Chaperone for the Mammalian Ribosome. *Molecular and Cellular Biology*, *28*(23), 7050–7065. https://doi.org/10.1128/mcb.01548-07

Majewski, L., Nowak, J., Sobczak, M., Karatsai, O., Havrylov, S., Lenartowski, R., Suszek, M., Lenartowska, M., & Redowicz, M. J. (2018). Myosin VI in the nucleus of neurosecretory PC12 cells: Stimulation-dependent nuclear translocation and interaction with nuclear proteins. *Nucleus*, *9*(1), 125–141. https://doi.org/10.1080/19491034.2017.1421881

Malgulwar, P. B., Danussi, C., Dharmaiah, S., Johnson, W., Singh, A., Rai, K., Rao, A., Huse, J. T. (2024). Sirtuin 2 inhibition modulates chromatin landscapes genome-wide to induce senescence in ATRX-deficient malignant glioma. *Neuro Oncol*. Jan 5;26(1):55-67. doi: 10.1093/neuonc/noad155. PMID: 37625115; PMCID: PMC10769000.

Marano, D., Fioriniello, S., Fiorillo, F., Gibbons, R. J., D'esposito, M., & Ragione, F. Della. (2019). ATRX contributes to MECP2-mediated pericentric heterochromatin organization during neural differentiation. *International Journal of Molecular Sciences*, *20*(21), 1–28. https://doi.org/10.3390/ijms20215371

Marquez-Lona, E. M., Tan, Z., & Schreiber, S. S. (2012). Nucleolar stress characterized by downregulation of nucleophosmin: A novel cause of neuronal degeneration. *Biochemical and Biophysical Research Communications*, *417*(1), 514–520. https://doi.org/10.1016/j.bbrc.2011.11.152

Matthews, D. A. (2001). Adenovirus Protein V Induces Redistribution of Nucleolin and B23 from Nucleolus to Cytoplasm. *Journal of Virology*, *75*(2), 1031–1038. https://doi.org/10.1128/jvi.75.2.1031-1038.2001

Maze, I., Wenderski, W., Noh, K. M., Bagot, R. C., Tzavaras, N., Purushothaman, I., Elsässer, S. J., Guo, Y., Ionete, C., Hurd, Y. L., Tamminga, C. A., Halene, T., Farrelly, L., Soshnev, A. A., Wen, D., Rafii, S., Birtwistle, M. R., Akbarian, S., Buchholz, B. A., Allis, C. D. (2015). Critical Role of Histone Turnover in Neuronal Transcription and Plasticity. *Neuron*, *87*(1), 77–94. https://doi.org/10.1016/j.neuron.2015.06.014

McDowell, T. L., Gibbons, R. J., Sutherland, H., O'Rourke, D. M., Bickmore, W. A., Pombo, A., Turley, H., Gatter, K., Picketts, D. J., Buckle, V. J., Chapman, L., Rhodes, D., & Higgs, D. R. (1999). Localization of a putative transcriptional regulator (ATRX) at pericentromeric heterochromatin and the short arms of acrocentric chromosomes. *Proceedings of the National Academy of Sciences of the United States of America*, *96*(24), 13983–13988. https://doi.org/10.1073/pnas.96.24.13983

McStay, B., & Grummt, I. (2008). The epigenetics of rRNA genes: From molecular to chromosome biology. *Annual Review of Cell and Developmental Biology*, *24*, 131–157. https://doi.org/10.1146/annurev.cellbio.24.110707.175259

Medrano-Fernández, A., & Barco, A. (2016). Nuclear organization and 3D chromatin architecture in cognition and neuropsychiatric disorders. *Molecular Brain*, *9*(1), 1–12. https://doi.org/10.1186/s13041-016-0263-x

Mélèse, T., & Xue, Z. (1995). The nucleolus: an organelle formed by the act of building a ribosome. *Current Opinion in Cell Biology*, *7*(3), 319–324. https://doi.org/10.1016/0955-0674(95)80085-9

Michaluk, P., Wawrzyniak, M., A lot, P., Szczot, M., Wyrembek, P., Mercik, K., Medvedev, N., Wilczek, E., De Roo, M., Zuschratter, W., Muller, D., Wilczynski, G.M., Mozrzymas, J.W., Stewart, M.G., Kaczmarek, L., Wlodarczyk, J. (2011). Influence of matrix metalloproteinase MMP-9 on dendritic spine morphology. *J Cell Sci.* 2011 Oct 1;124(Pt 19):3369-80. doi: 10.1242/jcs.090852. Epub 2011 Sep 6. PMID: 21896646.

Mierzejewski, P., & Kostowski, W. (2002). Rola hipokampa w patogenezie uzależnień i działaniu pozytywnie wzmacniającym substancji psychoaktywnych. *Alkoholizm i Narkomania*, *15*(2), 207-219.

Mifsud, B., Tavares-Cadete, F., Young, A. N., Sugar, R., Schoenfelder, S., Ferreira, L., Wingett, S. W., Andrews, S., Grey, W., Ewels, P. A., Herman, B., Happe, S., Higgs, A., Leproust, E., Follows, G. A., Fraser, P., Luscombe, N. M., & Osborne, C. S. (2015). Mapping long-range promoter contacts in human cells with high-resolution capture Hi-C. *Nature Genetics*, *47*(6), 598–606. https://doi.org/10.1038/ng.3286

Miller, O. L., & Beatty, B. R. (1969). Visualization of nucleolar genes. *Science*, *164*(3882), 955–957. https://doi.org/10.1126/science.164.3882.955

Misteli, T. (2020) The Self-Organizing Genome: Principles of Genome Architecture and Function. *Cell.* 2020 Oct 1;183(1):28-45. doi: 10.1016/j.cell.2020.09.014. Epub 2020 Sep 24. PMID: 32976797; PMCID: PMC7541718.

Monneron, A., Bernhard, W. (1969). Fine structural organization of the interphase nucleus in some mammalian cells. *J Ultrastruct Res.* May;27(3):266-88. doi: 10.1016/s0022-5320(69)80017-1. PMID: 5813971.

Montgomery, T. H. (1898). Comparative cytological studies, with especial regard to the morphology of the nucleolus. *Journal of Morphology*.

Morgan, A., Gandin, I., Belcaro, C., Palumbo, P., Palumbo, O., Biamino, E., Dal Col, V., Laurini, E., Pricl, S., Bosco, P., Carella, M., Ferrero, G. B., Romano, C., d'Adamo, A. P., Faletra, F., & Vozzi, D. (2015). Target sequencing approach intended to discover new mutations in non-syndromic intellectual disability. *Mutation Research - Fundamental and Molecular Mechanisms of Mutagenesis*, *781*, 32–36. https://doi.org/10.1016/j.mrfmmm.2015.09.002

Nazer, E. (2022). To be or not be (in the LAD): emerging roles of lamin proteins in transcriptional regulation. *Biochemical Society Transactions*, *50*(2), 1035–1044. https://doi.org/10.1042/BST20210858

Noh, K. M., Maze, I., Zhao, D., Xiang, B., Wenderski, W., Lewis, P. W., Shen, L., Li, H., & Allis, C. D. (2015). ATRX tolerates activity-dependent histone H3 methyl/phos switching to maintain repetitive element silencing in neurons. *Proceedings of the National Academy of Sciences of the United States of America*, *112*(22), 6820–6827. https://doi.org/10.1073/pnas.1411258112

Núñez Villacís, L., Wong, M. S., Ferguson, L. L., Hein, N., George, A. J., & Hannan, K. M. (2018). New Roles for the Nucleolus in Health and Disease. *BioEssays*, *40*(5), 8–10. https://doi.org/10.1002/bies.201700233

Olausson, K. H., Nistér, M., & Lindström, M. S. (2012). P53 -Dependent and -Independent Nucleolar Stress Responses. *Cells*, *1*(4), 774–798. https://doi.org/10.3390/cells1040774

Otmakhov, N., Khibnik, L., Otmakhova, N., Carpenter, S., Riahi, S., Asrican, B., & Lisman, J. (2004).

Forskolin-Induced LTP in the CA1 Hippocampal Region Is NMDA Receptor Dependent. *Journal of Neurophysiology*, *91*(5), 1955–1962. https://doi.org/10.1152/jn.00941.2003

Pang, Y., Chen, X., Ji, T., Cheng, M., Wang, R., Zhang, C., Liu, M., Zhang, J., & Zhong, C. (2023). The Chromatin Remodeler ATRX: Role and Mechanism in Biology and Cancer. *Cancers*, *15*(8). https://doi.org/10.3390/cancers15082228

Percy, A. K., Neul, J. L., Benke, T. A., Marsh, E. D., & Glaze, D. G. (2023). A review of the Rett Syndrome Behaviour Questionnaire and its utilization in the assessment of symptoms associated with Rett syndrome. *Frontiers in Pediatrics*, *11*(July), 1–9. https://doi.org/10.3389/fped.2023.1229553

Perry, R. P. (1967). The Nucleolus and the Synthesis of Ribosomes. *Progress in Nucleic Acid Research and Molecular Biology*, *6*(C), 219–257. https://doi.org/10.1016/S0079-6603(08)60528-0

Picketts, D. J., Higgs, D. R., Bachoo, S., Blake, D. J., Quarrell, O. W. J., & Gibbons, R. J. (1996). ATRX encodes a novel member of the SNF2 family of proteins: Mutations point to a common mechanism underlying the ATR-X syndrome. *Human Molecular Genetics*, *5*(12), 1899–1907. https://doi.org/10.1093/hmg/5.12.1899

Picketts, D. J., Tastan, A. O., Higgs, D. R., & Gibbons, R. J. (1998). Comparison of the human and murine ATRX gene identifies highly conserved, functionally important domains. *Mammalian Genome*, *9*(5), 400–403. https://doi.org/10.1007/s003359900781

Placek, B. J., & Gloss, L. M. (2002). The N-terminal tails of the H2A-H2B histones affect dimer structure and stability. *Biochemistry*, *41*(50), 14960–14968. https://doi.org/10.1021/bi026283k

Ratnakumar, K., & Bernstein, E. (2013). ATRX: The case of a peculiar chromatin remodeler. *Epigenetics*, *8*(1), 3–9. https://doi.org/10.4161/epi.23271

Renner, J. & Rasia-Filho, A. A. (2023). Morphological Features of Human Dendritic Spines. *Adv Neurobiol.* 2023;34:367-496. doi: 10.1007/978-3-031-36159-3_9. PMID: 37962801.

Reymann, K. G., & Frey, J. U. (2007). The late maintenance of hippocampal LTP: Requirements, phases, „synaptic tagging", „late-associativity" and implications. *Neuropharmacology*, *52*(1), 24–40. https://doi.org/10.1016/j.neuropharm.2006.07.026

Richter, J. D., & Klann, E. (2009). Making synaptic plasticity and memory last: Mechanisms of translational regulation. *Genes and Development*, *23*(1), 1–11. https://doi.org/10.1101/gad.1735809

Rodrigues, S. M., Schafe, G. E., & Ledoux, J. E. (2004). Molecular mechanisms underlying emotional learning and memory in the lateral amygdala. *Neuron*, *44*(1), 75–91. https://doi.org/10.1016/j.neuron.2004.09.014

Rorbach-Dolata, A., Kubis, A., & Piwowar, A. (2017). Epigenetic modifications: An important mechanism in diabetic disturbances. *Postępy Higieny i Medycyny Doświadczalnej*, *71*(0), 0–0. https://doi.org/10.5604/01.3001.0010.6156

Roszkowska, M., Skupien, A., Wójtowicz, T., Konopka, A., Gorlewicz, A., Kisiel, M., Bekisz, M., Ruszczycki, B., Dolezyczek, H., Rejmak, E., Knapska, E., Mozrzymas, J. W., Wlodarczyk, J., Wilczynski, G. M., & Dzwonek, J. (2016). CD44: A novel synaptic cell adhesion molecule regulating structural and functional plasticity of dendritic spines. *Molecular Biology of the Cell*, *27*(25), 4055–4066. https://doi.org/10.1091/mbc.E16-06-0423

Rubbi, C. P., & Milner, J. (2003). Disruption of the nucleolus mediates stabilization of p53 in response to DNA damage and other stresses. *The EMBO Journal*, *22*(22), 6068–6077.

Ryan, D. P., & Owen-hughes, T. (2014). *Europe PMC Funders Group Snf2-family proteins : chromatin remodellers for any occasion*. *15*(5), 649–656. https://doi.org/10.1016/j.cbpa.2011.07.022.Snf2-family

Sacharowski, S. P. & Sarnowski, T. J. (2019). Mechanizmy kontrolujące strukturę chromatyny. *Postepy Biochem*. Mar 22;65(1):9-20. doi: 10.18388/pb.2019_252.

Seah, C., Levy, M. A., Jiang, Y., Mokhtarzada, S., Higgs, D. R., Gibbons, R. J., & Bérubé, N. G. (2008). Neuronal death resulting from targeted disruption of the Snf2 protein ATRX is mediated by p53. *Journal of Neuroscience*, *28*(47), 12570–12580. https://doi.org/10.1523/JNEUROSCI.4048-08.2008

Selakovic, D., Joksimovic, J., Jovicic, N., Mitrovic, S., Mihailovic, V., Katanic, J., Milovanovic, D., Pantovic, S., Mijailovic, N., & Rosic, G. (2019). The impact of hippocampal sex hormones receptors in modulation of depressive-like behavior following chronic anabolic androgenic steroids and exercise protocols in rats. *Frontiers in Behavioral Neuroscience*, *13*(February), 1–15. https://doi.org/10.3389/fnbeh.2019.00019

Sharifi, S. & Bierhoff H. (2018). Regulation of RNA Polymerase I Transcription in Development, Disease, and Aging. *Annu Rev Biochem.* 2018 Jun 20;87:51-73. doi: 10.1146/annurev-biochem-062917-012612. Epub 2018 Mar 28. PMID: 29589958.

Shioda, N., Yabuki, Y., Yamaguchi, K., Onozato, M., Li, Y., Kurosawa, K., Tanabe, H., Okamoto, N., Era, T., Sugiyama, H., Wada, T., Fukunaga K. (2018).Targeting G-quadruplex DNA as cognitive function therapy for ATR-X syndrome. *Nat Med.* 2018 Jun;24(6):802-813. doi: 10.1038/s41591-018-0018-6.

Sirri, V., Urcuqui-Inchima, S., Roussel, P., Hernandez-Verdun, D. Nucleolus: the fascinating nuclear body. (2208). *Histochem Cell Biol*. 2008 Jan;129(1):13-31. doi: 10.1007/s00418-007-0359-6. Epub 2007 Nov 29. PMID: 18046571; PMCID: PMC2137947.

Skupien, A., Konopka, A., Trzaskoma, P., Labus, J., Gorlewicz, A., Swiech, L., Babraj, M., Dolezyczek, H., Figiel, I., Ponimaskin, E., Wlodarczyk, J., Jaworski, J., Wilczynski, G. M., Dzwonek, J. (2014). CD44 regulates dendrite morphogenesis through Src tyrosine kinase-dependent positioning of the Golgi. *J Cell Sci.* 2014 Dec 1;127(Pt 23):5038-51. doi: 10.1242/jcs.154542. Epub 2014 Oct 9. PMID: 25300795.

Skupien-Jaroszek, A., Walczak, A., Czaban, I., Pels, K. K., Szczepankiewicz, A. A., Krawczyk, K., Ruszczycki, B., Wilczynski, G. M., Dzwonek, J., Magalska, A. (2021). The interplay of seizures-induced axonal sprouting and transcription-dependent Bdnf repositioning in the model of temporal lobe epilepsy. *PLoS One*. 2021 Jun 4;16(6):e0239111. doi: 10.1371/journal.pone.0239111. PMID: 34086671; PMCID: PMC8177504.

Spector, D. L. Nuclear domains. (2001) *J Cell Sci.* Aug;114(Pt 16):2891-3. doi:10.1242/jcs.114.16.2891.

Stoppini, L., Buchs, P. A., & Muller, D. (1991). A simple method for organotypic cultures of nervous tissue. *Journal of Neuroscience Methods*, *37*(2), 173–182. https://doi.org/10.1016/0165-0270(91)90128-M

Talbert, P. B., & Henikoff, S. (2010). Histone variants ancient wrap artists of the epigenome. *Nature Reviews Molecular Cell Biology*, *11*(4), 264–275. https://doi.org/10.1038/nrm2861

Tao-Cheng, J. H.(2019) Stimulation induces gradual increases in the thickness and curvature of postsynaptic density of hippocampal CA1 neurons in slice cultures. *Mol Brain.* 12, 44. https://doi.org/10.1186/s13041-019-0468-x

Thielmann, H. W., Popanda, O., & Staab, H. J. (1999). Subnuclear distribution of DNA topoisomerase I and Bax protein in normal and xeroderma pigmentosum fibroblasts after irradiation with UV light and γ rays or treatment with topotecan. *Journal of Cancer Research and Clinical Oncology*, *125*(3–4), 193–208. https://doi.org/10.1007/s004320050263

Trovato, M., Patil, V., Gehre, M., & Noh, K. M. (2020). Histone Variant H3.3 Mutations in Defining the Chromatin Function in Mammals. *Cells*, *9*(12). https://doi.org/10.3390/cells9122716

Truch, J., Downes, D. J., Scott, C., Gür, E. R., Telenius, J. M., Repapi, E., Schwessinger, R., Gosden, M., Brown, J. M., Taylor, S., Cheong, P. L., Hughes, J. R., Higgs, D. R., & Gibbons, R. J. (2022). The chromatin remodeller ATRX facilitates diverse nuclear processes, in a stochastic manner, in both heterochromatin and euchromatin. *Nature Communications*, *13*(1), 1–16. https://doi.org/10.1038/s41467-022-31194-7

Tsekrekou, M., Stratigi, K., & Chatzinikolaou, G. (2017). The nucleolus: In genome maintenance and repair. *International Journal of Molecular Sciences*, *18*(7). https://doi.org/10.3390/ijms18071411

Udugama, M., Sanij, E., Voon, H. P. J., Son, J., Hii, L., Henson, J. D., Chan, F. L., Chang, F. T. M., Liu, Y., Pearson, R. B., Kalitsis, P., Mann, J. R., Collas, P., Hannan, R. D., Wong, L. H. (2018). Ribosomal DNA copy loss and repeat instability in ATRX-mutated cancers. *Proc Natl Acad Sci U S A*. 2018 May 1;115(18):4737-4742. doi: 10.1073/pnas.1720391115. Epub 2018 Apr 18. PMID: 29669917; PMCID: PMC5939086.

Urbanska, M., Blazejczyk, M., Jaworski, J. (2008). Molecular basis of dendritic arborization. *Acta Neurobiol Exp* (Wars). 2008;68(2):264-88. doi: 10.55782/ane-2008-1695. PMID: 18511961.

Urbanska, M., Swiech, L., Jaworski, J. (2012). Developmental plasticity of the dendritic compartment: focus on the cytoskeleton. *Adv Exp Med Biol.* 2012;970:265-84. doi: 10.1007/978-3-7091-0932-8_12. PMID: 22351060.

Vargas, J. P., Rodríguez, F., López, J. C., Arias, J. L., & Salas, C. (2000). Spatial learning-induced increase in the argyrophilic nucleolar organizer region of dorsolateral telencephalic neurons in goldfish. *Brain Research*, *865*(1), 77–84. https://doi.org/10.1016/S0006-8993(00)02220-4

Villard, L., Lossi, A. M., Cardoso, C., Proud, V., Chiaroni, P., Colleaux, L., Schwartz, C., & Fontés, M. (1997). Determination of the genomic structure of the XNP/ATRX gene encoding a potential zinc finger helicase. *Genomics*, *43*(2), 149–155. https://doi.org/10.1006/geno.1997.4793

Vincent, J. A., Kwong, T. J., & Tsukiyama, T. (2008). ATP-dependent chromatin remodeling shapes the DNA replication landscape. *Nature Structural and Molecular Biology*, *15*(5), 477–484. https://doi.org/10.1038/nsmb.1419

Wada, T., Ban, H., Matsufuji, M., Okamoto, N., Enomoto, K., Kurosawa, K., Aida, N. (2013). Neuroradiologic features in X-linked α-thalassemia/mental retardation syndrome. *AJNR Am J Neuroradiol.* 2013 Oct;34(10):2034-8. doi: 10.3174/ajnr.A3560.

Walczak, A., Szczepankiewicz, A. A., Ruszczycki, B., Magalska, A., Zamlynska, K., Dzwonek, J., Wilczek, E., Zybura-Broda, K., Rylski, M., Malinowska, M., Dabrowski, M., Szczepinska, T., Pawlowski, K., Pyskaty, M., Wlodarczyk, J., Szczerbal, I., Switonski, M., Cremer, M., & Wilczynski, G. M. (2013). Novel higher-order epigenetic regulation of the Bdnf gene upon seizures. *Journal of Neuroscience*, *33*(6), 2507–2511. https://doi.org/10.1523/JNEUROSCI.1085-12.2013

Warmerdam, D. O., & Wolthuis, R. M. F. (2019). Keeping ribosomal DNA intact: a repeating challenge. *Chromosome Research*, *27*(1–2), 57–72. https://doi.org/10.1007/s10577-018-9594-z

Wenderski, W. & Maze, I. (2016). Histone turnover and chromatin accessibility: Critical mediators of neurological development, plasticity, and disease. *Bioessays*. 2016 May;38(5):410-9. doi: 10.1002/bies.201500171. Epub 2016 Mar 15. PMID: 26990528; PMCID: PMC4968875.

Wilczynski, G. M. (2014). Significance of higher-order chromatin architecture for neuronal function and dysfunction. *Neuropharmacology*, *80*, 28–33. https://doi.org/10.1016/J.NEUROPHARM.2014.01.016

Wong, L. H., McGhie, J. D., Sim, M., Anderson, M. A., Ahn, S., Hannan, R. D., George, A. J., Morgan, K. A., Mann, J. R., & Choo, K. H. A. (2010). ATRX interacts with H3.3 in maintaining telomere structural integrity in pluripotent embryonic stem cells. *Genome Research*, *20*(3), 351–360. https://doi.org/10.1101/gr.101477.109

Wu, H., & Zhang, Y. (2014). Reversing DNA methylation: Mechanisms, genomics, and biological functions. *Cell*, *156*(1–2), 45–68. https://doi.org/10.1016/j.cell.2013.12.019

Xue, Y., Gibbons, R., Yan, Z., Yang, D., McDowell, T. L., Sechi, S., Qin, J., Zhou, S., Higgs, D., & Wang, W. (2003). The ATRX syndrome protein forms a chromatin-remodeling complex with Daxx and localizes in promyelocytic leukemia nuclear bodies. *Proceedings of the National Academy of Sciences of the United States of America*, *100*(19), 10635–10640. https://doi.org/10.1073/pnas.1937626100

Yamada, S. & Nelson, W. J. (2007). Synapses: sites of cell recognition, adhesion, and functional specification. *Annu Rev Biochem.* 2007;76:267-94. doi: 10.1146/annurev.biochem.75.103004.142811. PMID: 17506641; PMCID: PMC3368613.

Yang, K., Wang, M., Zhao, Y., Sun, X., Yang, Y., Li, X., Zhou, A., Chu, H., Zhou, H., Xu, J., Wu, M., Yang, J., & Yi, J. (2016). A redox mechanism underlying nucleolar stress sensing by nucleophosmin. *Nature Communications*, *7*, 1–16. https://doi.org/10.1038/ncomms13599

Yang, X., Khosravi-Far, R., Chang, H. Y., Baltimore, D. (1997). Daxx, a novel Fas-binding protein that activates JNK and apoptosis. *Cell*. Jun 27;89(7):1067-76. doi: 10.1016/s0092-8674(00)80294-9. PMID: 9215629; PMCID: PMC2989411.

Yap, E. L., & Greenberg, M. E. (2018). Activity-regulated transcription: Bridging the gap between neural activity and behavior. *Neuron*, *100*(2), 330. https://doi.org/10.1016/J.NEURON.2018.10.013

Yasuzumi, G., Sawada, T., Sugihara, R., Kiriyama, M., & Sugioka, M. (1958). Electron microscope researches on the ultrastructure of nucleoli in animal tissues. *Zeitschrift für Zellforschung und Mikroskopische Anatomie*, *48*(1), 10–23. https://doi.org/10.1007/BF00496710

Yogev, O., Saadon, K., Anzi, S., Inoue, K., & Shaulian, E. (2008). DNA damage-dependent translocation of B23 and p19ARF is regulated by the Jun N-terminal kinase pathway. *Cancer Research*, *68*(5), 1398–1406. https://doi.org/10.1158/0008-5472.CAN-07-2865

Yung, B. Y. M., Busch, H., & Chan, P. K. (1985). Translocation of nucleolar phosphoprotein B23 (37kDa pI 5.1) induced by selective inhibitors of ribosome synthesis. *BBA - Gene Structure and Expression*, *826*(4), 167–173. https://doi.org/10.1016/0167-4781(85)90002-8